



Instituto de
Ciencias de la
Vid y del Vino



CSIC



Status and control of grapevine trunk diseases at the nursery and young vineyard level

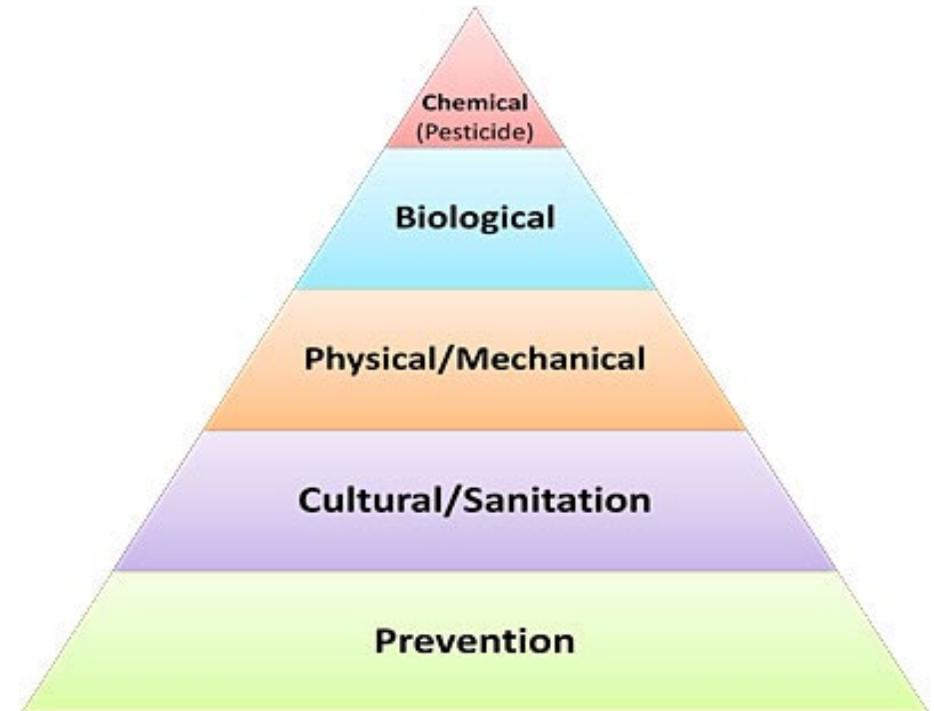
David Gramaje

GRAPEVINE TRUNK DISEASES (GTD)

Complexity of this pathosystem

- 1 Many fungal species associated with GTD symptoms
- 2 Fungi with different biology and epidemiology
- 3 No curative measures are known for control of GTD

INTEGRATED DISEASE MANAGEMENT STRATEGY



Integrated disease management strategy

Nursery mother blocks



Propagation processes



Nursery field



Newly established vineyards



Integrated disease management strategy

Nursery mother blocks

Cultural practices and sanitation

Pruning wound protection

Propagation processes

Chemical/biological/physical treatments

Nursery field

Biofumigation

Newly established vineyards

Selection of planting material

Site preparation and vine management

Biological control

Pruning wound protection

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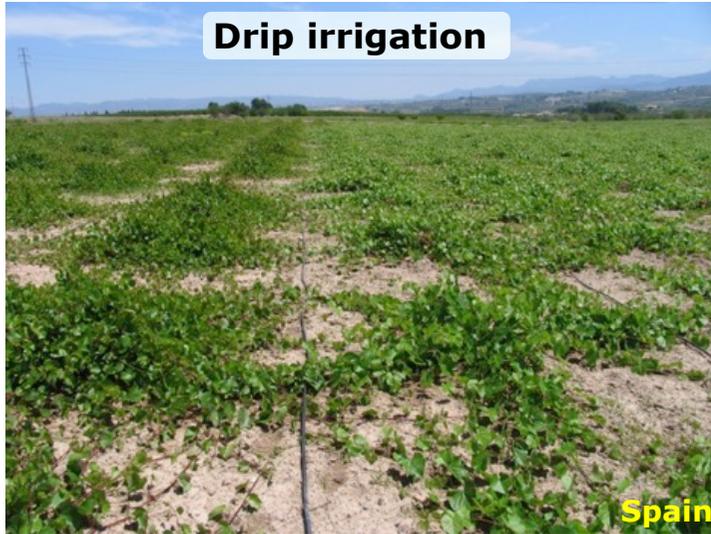
Nursery mother blocks

Cultural practices and sanitation: rootstock mother vine cultivation



Nursery mother blocks

Cultural practices and sanitation: irrigation



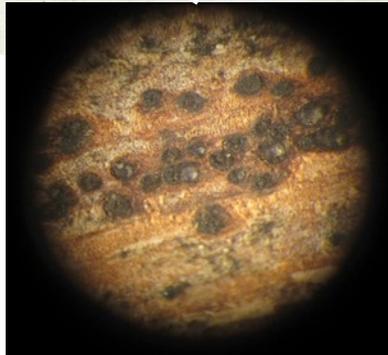
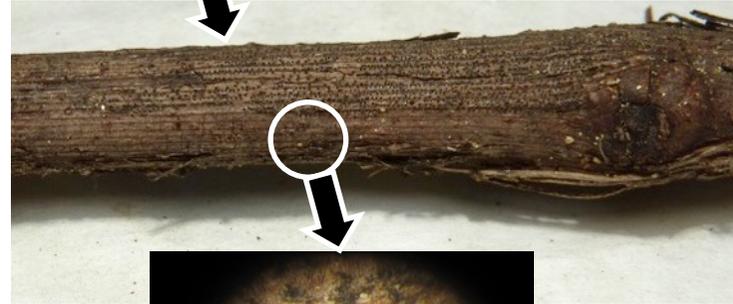
Botryosphaeriaceae spp.
Úrbez-Torres et al. 2010

Phaeoacremonium spp.
Gubler et al. 2013

Diatrypaceae spp.
Úrbez-Torres et al. 2019

Nursery mother blocks & vineyard

Cultural practices and sanitation: **removal of dead wood or pruning debris**



Nursery mother blocks & vineyard

Cultural practices and sanitation: removal of dead wood or pruning debris

1) Composting



lifesarmiento.eu



Lecomte et al. 2006 *Phytopathol. Mediterr* 45

2) Mulching



A protocol for the management of grapevine rootstock mother vines to reduce latent infections by grapevine trunk pathogens in cuttings

HELEN WAITE¹, JOSEP ARMENGOL², REGINA BILLONES-BAAIJENS³, DAVID GRAMAJE⁴, FRANCOIS HALLEEN^{5,6}, STEFANO DI MARCO⁷ and RICHARD SMART⁸

Phytopathologia Mediterranea (2018), 57, 3, 384–398

- ✓ Training systems
- ✓ Irrigation
- ✓ Cultural practices and sanitation
- ✓ Pruning wound protection

3) Burning



Winter pruning: practice and protection

Diatrypaceae spp.
Botryosphaeriaceae spp.
Basidiomycetes
Phaeomoniella chlamydospora
Phaeoacremonium spp.



Mother vine



Mature vine

Pruning wound protection

Mastic/paste + fungicides

Benzimidazole carbamate mode of action group (benomyl, carbendazim, and thiophanate methyl)

Eutypa dieback

Australia (Sosnowski et al. 2008 Aus. J. Grape Wine Res. 14)
USA (Rolshausen et al. 2010 Am. J. Enol. Vitic. 61)

Botryosphaeria dieback

South Africa (Halleen et al. 2010)
Chile (Díaz and Latorre 2013 Crop Prot. 46)
USA (Rolshausen et al. 2010 Am. J. Enol. Vitic. 61)

Esca

Chile (Díaz and Latorre 2013 Crop Prot. 46)
South Africa (Mutawila et al. 2015 BioControl 60)

The demethylation inhibitors (tebuconazole & flusilazole)

Eutypa dieback

Australia (Ayres et al. 2017 Aus. J. Grape Wine Res. 23)
South Africa (Halleen et al. 2010 S. Afr. J. Enol. Vitic. 31)

Botryosphaeria dieback

Chile (Díaz and Latorre 2013 Crop Prot. 46)
New Zealand (Amponsah et al. 2012 Pest. Manag. Sci. 68)
Australia (Pitt et al. 2012 Plant Dis. 96)
South Africa (Halleen et al. 2010 S. Afr. J. Enol. Vitic. 31)

**Restrictions and difficulties that
chemicals are facing in most
countries around the world**



**Mastic or
paste**

Pruning wound protection



Pruning-wound protectants for trunk-disease management in California table grapes

Albre A. Brown^a, Renaud Travadon^a, Daniel P. Lawrence^a, Gabriel Torres^b, George Zhuang^c, Kendra Baumgartner^{d,*}

Protection of grapevine pruning wounds against *Phaeomoniella chlamydospora* and *Diplodia seriata* by commercial biological and chemical methods

María del Pilar Martínez-Diz^{a,b}, Emilia Díaz-Losada^a, Ángela Díaz-Fernández^a, Yolanda Bouzas-Cid^a, David Gramaje^{c,*}

pyraclostrobin + boscalid
with or without a liquid polymer



ALTERNATIVE: Biological Control Agents (BCA)

Good performance with *Trichoderma* spp.:

South Africa:

Kotze et al. 2011 *Phytopathol. Mediterr.* 50

Mutawila et al. 2016 *Aus. J. Grape Wine Res.* 22

Italy:

Di Marco et al. 2004 *Phytopathol. Mediterr.* 43

Bad performance with *Trichoderma* spp.:

Spain:

Martínez-Diz et al. 2021 *Crop Protection* 143

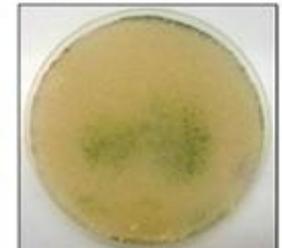
In general, BCA have shown variable results for preventing infection by GTD pathogens



Trichoderma harzianum



Trichoderma reesei



Trichoderma atroviride

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Selection of planting material

Site preparation and vine management

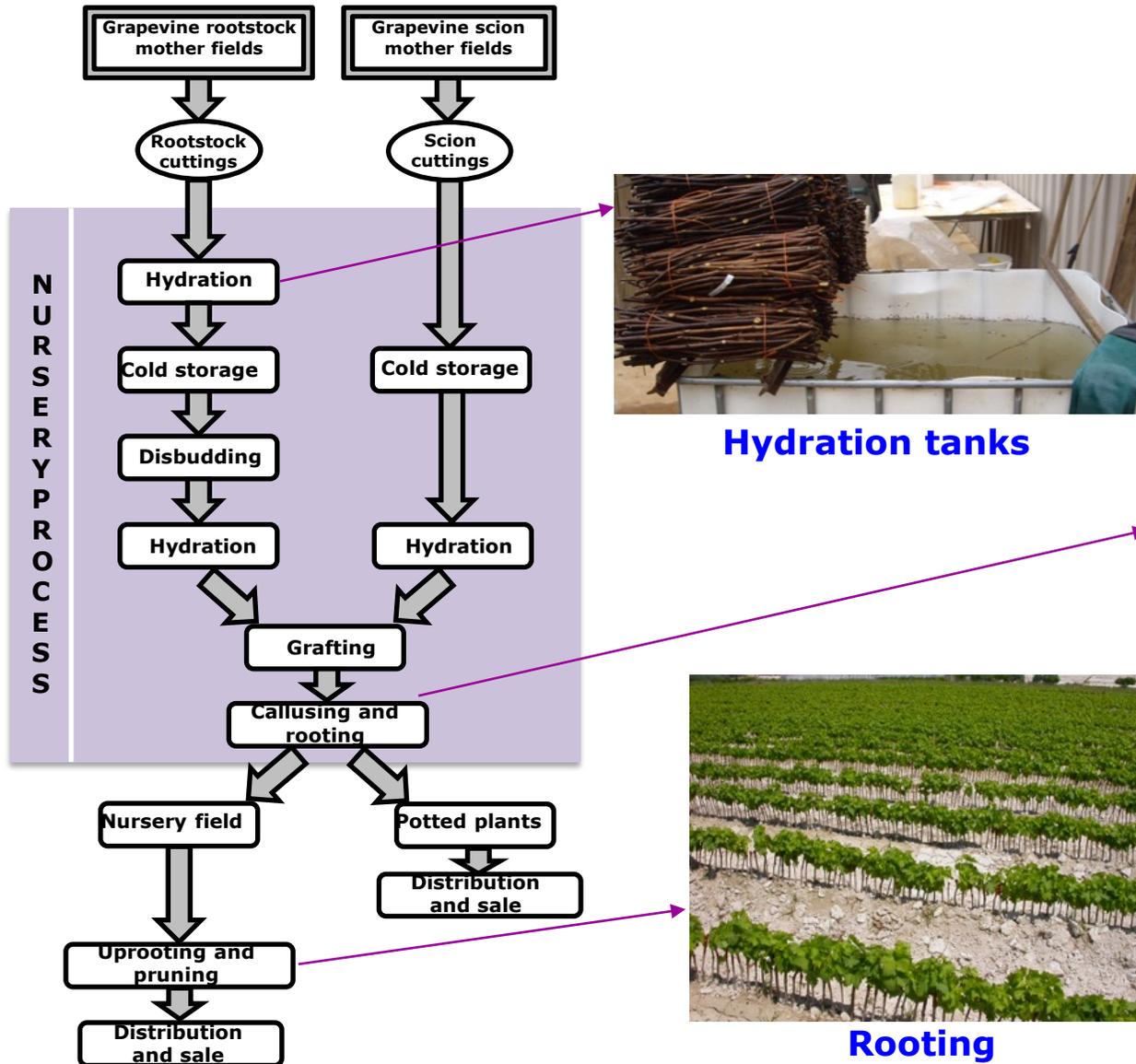
Biological control

Pruning wound protection

NURSERIES ARE SOURCES OF DISEASED MATERIAL

1 Nurseries are favorable for fungal trunk pathogens

au
wi
sp
su
au
wi
sp



- ✓ Wet and humid conditions
- ✓ High root density
- ✓ Close spacing of plants

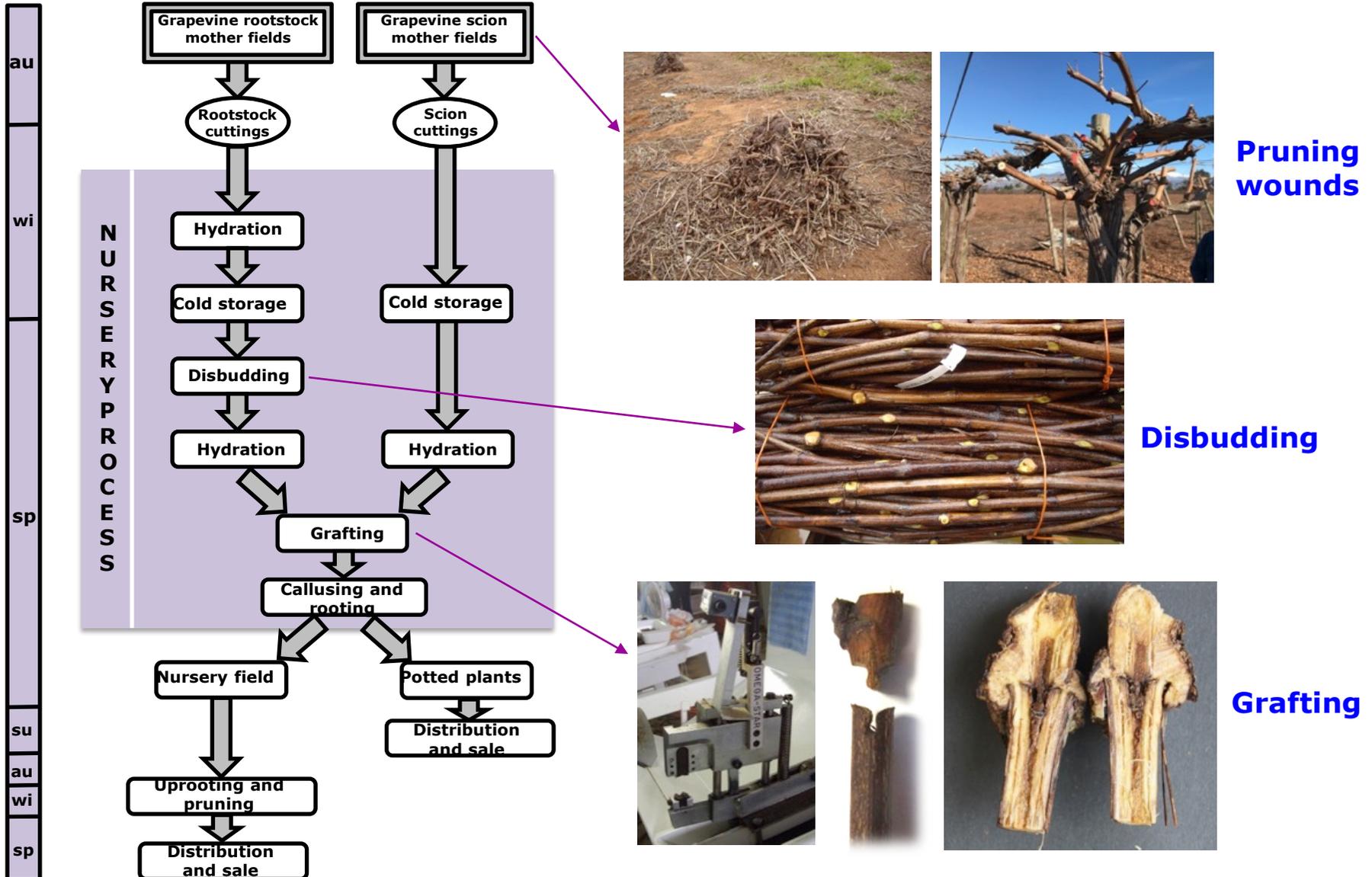
Hydration tanks

Callusing

Rooting

NURSERIES ARE SOURCES OF DISEASED MATERIAL

2 Practices increase infection risk



NURSERIES ARE SOURCES OF DISEASED MATERIAL

③ Diseased plants are difficult to detect

External symptomless plants



Latent pathogens: asymptomatic tissues



Pathogenic: biotic and/or abiotic stress factors

Destructive sampling

THE SOLUTION: CLEAN PLANT PRODUCTION

START CLEAN, KEEP IT CLEAN

- ✓ **Use pathogen-free plant material**
- ✓ **Clean grafting machines frequently**
- ✓ **Maintain a high standard of general cleanliness in the nursery, particularly the grafting room, callusing room and cool room**
- ✓ **Never reuse callusing media**
- ✓ **Avoid soaking cuttings**

Propagation processes in the nursery

CHEMICAL CONTROL

Carbendazim [Petri disease and Botryosphaeria dieback]

(Gramaje et al. 2009 Crop Prot. 28; Billones-Baaijens et al. 2015 Phytopathol. Mediterr. 54)

Cyprodinil + Fludioxinil (Switch®) [Black-foot disease and Botryosphaeria dieback]

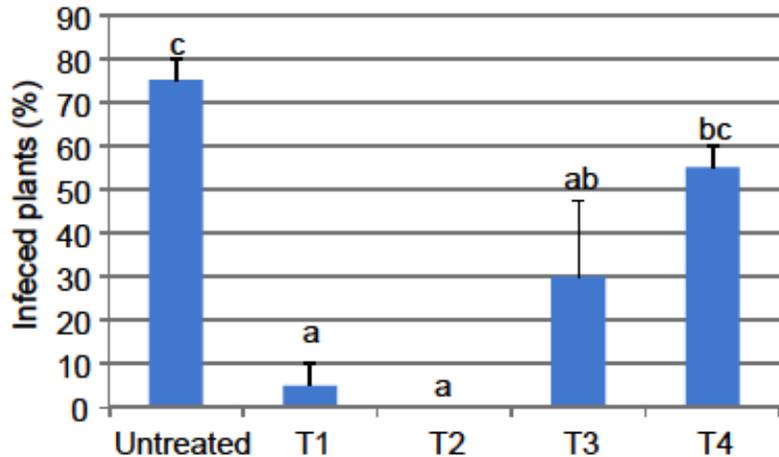
(Rego et al. 2009 Phytopathol. Mediter. 48)

FUNGICIDE AUTHORIZED IN EUROPE AGAINST GTDs IN NURSERIES: Chinosol (Hydroxyquinoline sulphate) in Germany

BIOLOGICAL CONTROL

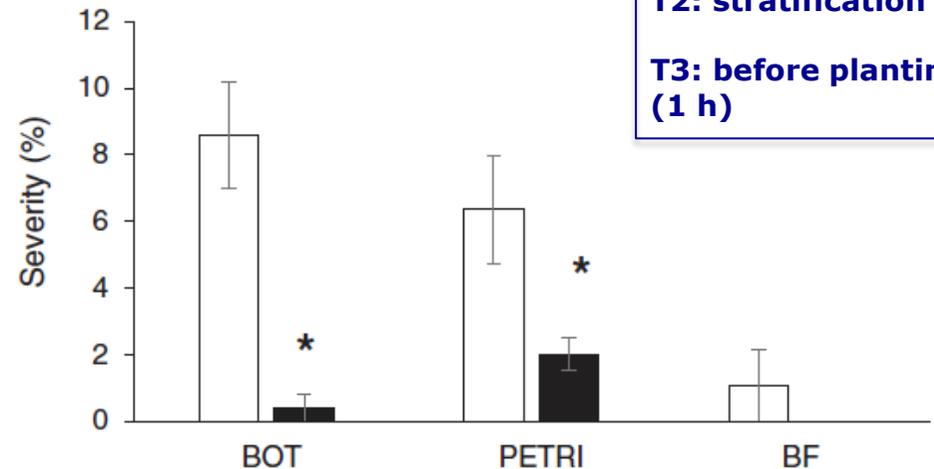
Trichoderma atroviride SC1 (Vintec®)

Phaeomoniella chlamydospora



Pertot et al. 2016. BioControl 61

T1: pre-storage hydration
T2: pre-grafting hydration



□ Control ■ Trichoderma SC1

Berbegal et al. 2019. Pest Manag. Sci.

T1: prior to grafting (24 h)

T2: stratification

T3: before planting (1 h)

Propagation processes in the nursery

HOT-WATER TREATMENT (HWT)

Standard treatment: 50°C – 30 min.

Several pests and diseases: Phytoplasma organisms. *Xylella fastidiosa* (50°C – 45 min. EFSA Journal 2015)

1 Some *Vitis vinifera* varieties are more sensitive to HWT than others (Waite et al. 2001)

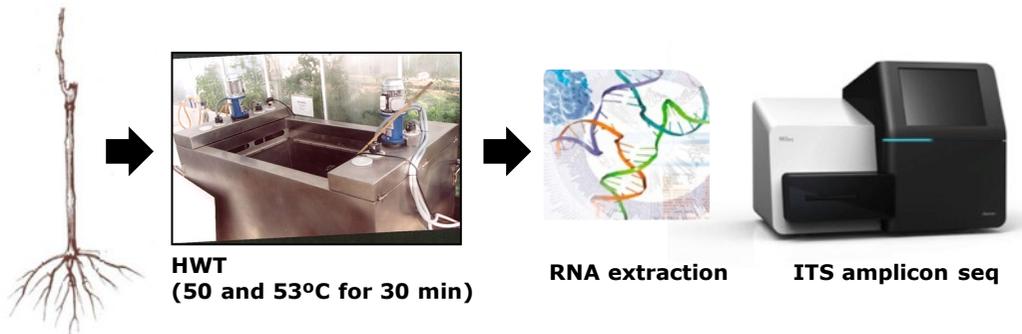
Pinot Noir: highly sensitive

Chardonnay, Reisling and Merlot: moderately sensitive

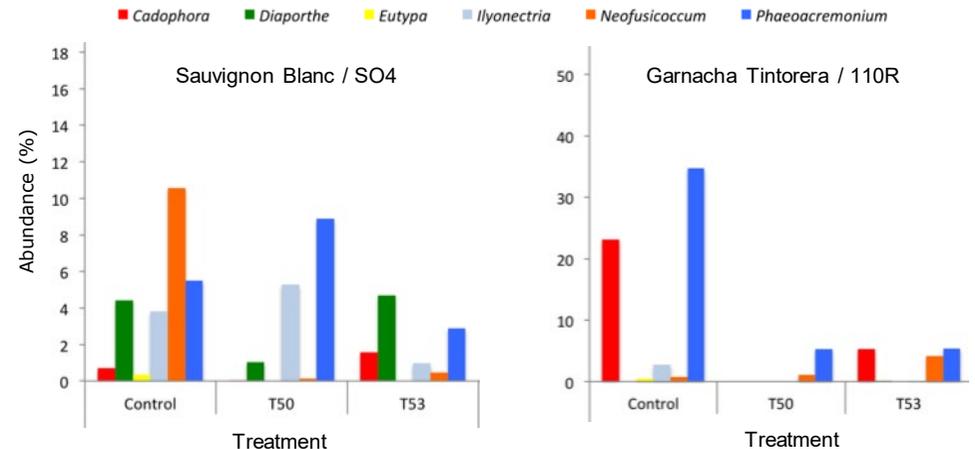
Cabernet Sauvignon: the least sensitive

2 Tolerance of plants to HWT is affected by the climate in which the cuttings are grown

3 HWT is not completely effective in eliminating fungal trunk disease pathogens growth



Eichmeier et al. 2018. Fungal Ecology 36



Integrated disease management strategy

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Nursery field

Biofumigation

Newly established vineyards

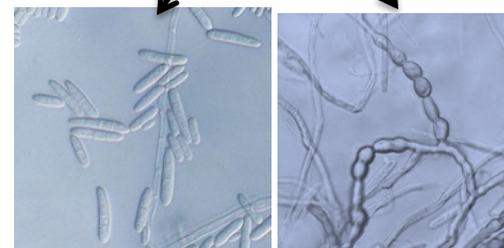
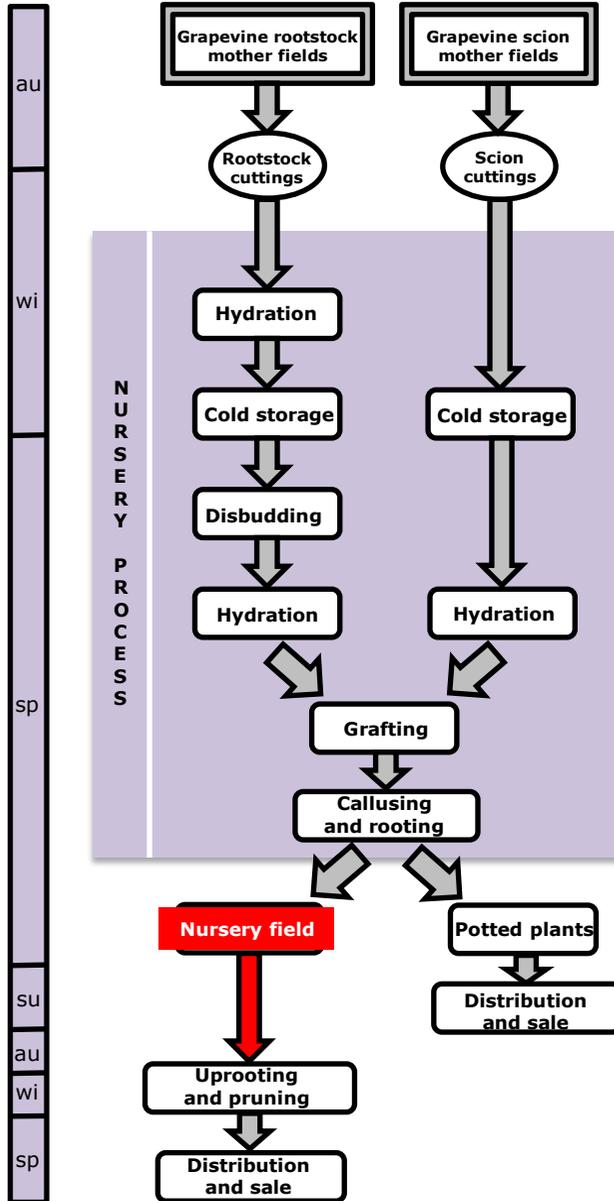
Selection of planting material

Site preparation and vine management

Biological control

Pruning wound protection

Nursery propagation beds



**BLACK-FOOT
DISEASE
PATHOGENS**

Nursery propagation beds

CROP ROTATION IN NURSERY FIELDS

Increase of black-foot disease incidence

Portugal

Grapevines + 3-year rotation (e.g. potato, garlic, carrot, cereals)

(Rego et al. 2009 *Phytopathol. Mediterr.* 48)

South Africa

Grapevines every second year + cover crop

(Halleen et al. 2003 *Australa. Plant Pathol.* 32)

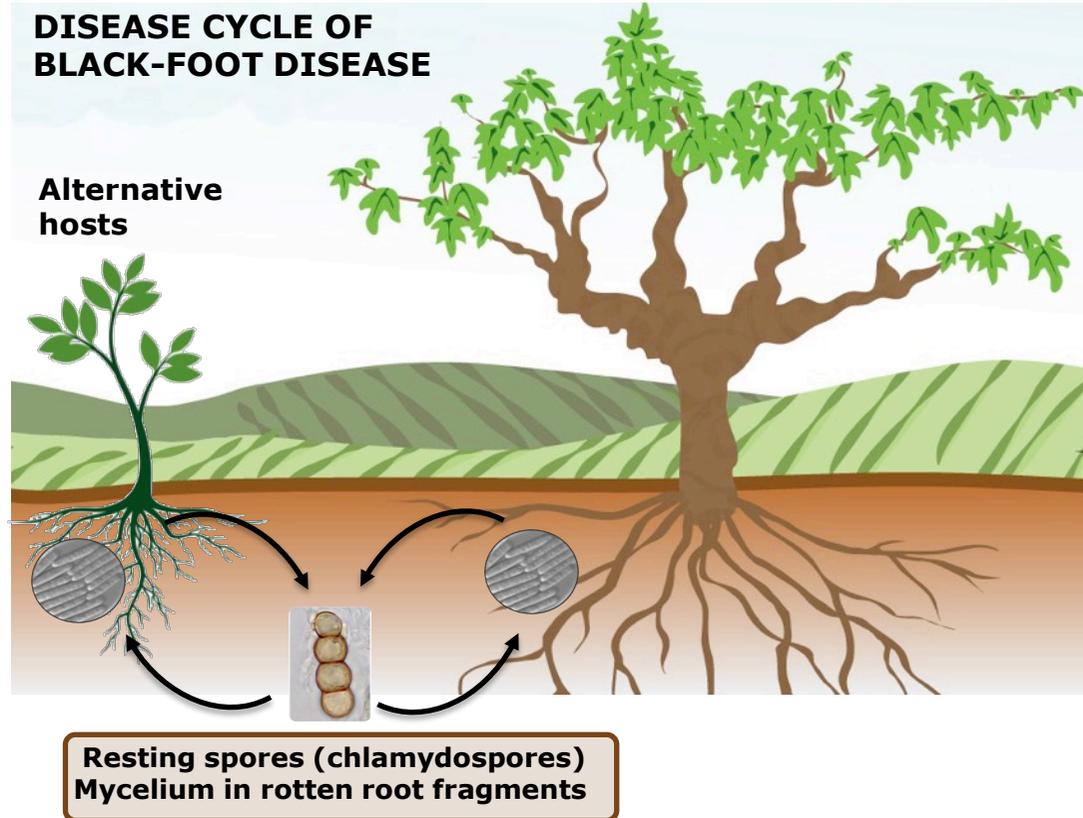
Fungal detection in soil during rotation

Wheat and barley

Portugal (Cardoso et al. 2013 *Phytop. Mediterr.* 52)

Spain (Berlanas et al. 2017 *Plant Soil* 417)

✓ Biofumigation



Indian mustard seed meal

New Zealand

Barbour et al. 2014 *Phytopathol. Mediterr.* 53
Bleach et al. 2010 *Phytopathol. Mediterr.* 49

White mustard

Spain

Berlanas et al. 2018 *Pest Manag. Sci.* 74

Canada

Richards et al. 2020 *Diversity* 12

Reduction of black-foot disease incidence and fungal inoculum in soil

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Commercial production settings

SELECTION OF GRAFTED PLANTS

- ✓ Vines should be of an even size and vigor
- ✓ Vines should be free of signs of disease
- ✓ Vines should not be broken or show signs of damage
- ✓ Each vine should have at least 1 well-developed shoot with healthy buds
- ✓ Each vine should have at least 3 healthy, undamaged, evenly spaced roots



Petri disease pathogens

Commercial production settings

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Grafts should be fully healed, not overgrown and not able to be broken by "moderate" pressure applied by the thumb

Commercial production settings

SITE PREPARATION FOR NEWLY ESTABLISHED VINEYARDS

Preplanting method: mustard seed meal can be incorporated into the soil, or a rotation crop of mustard can be grown until flowering and then incorporated before planting

Planting practice: compacted soil layers should be broken up before planting and vine should be well placed in large planting holes. Avoid soil compaction with heavy vehicles.



"J-rooting"



Photo: J. Vivancos (APA Québec)

Commercial production settings

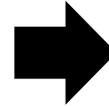
PREPLANTING TREATMENTS: Biological control agents

Pest Management
Science



Investigation of *Trichoderma* species colonization of nursery grapevines for improved management of black foot disease

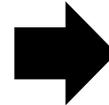
Wynand J van Jaarsveld,^{a,b} Francois Halleen,^{a,b} Michael C Bester,^a Romain JG Pierron,^c Elodie Stempien^a and Lizel Mostert^{a*}



Trichoderma spp. were not sufficient to prevent infections by BFD pathogens, but a certain degree of protection was obtained in the basal ends.

Field evaluation of biocontrol agents against black-foot and Petri diseases of grapevine

María del Pilar Martínez-Diz,^{a,b} Emilia Díaz-Losada,^a Marcos Andrés-Sodupe,^c Rebeca Bujanda,^c María M Maldonado-González,^c Sonia Ojeda,^c Amira Yacoub,^d Patrice Rey^d and David Gramaje^{c*}



The combination of the disease-suppressive activity of two or more beneficial microbes is required to prevent fungal infection

Streptomyces sp. E1 + R4
Pythium oligandrum Po37
Trichoderma atroviride SC1
Trichoderma koningii TK7
Pseudomonas fluorescens + *Bacillus atrophaeus*



Commercial production settings

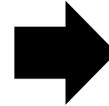
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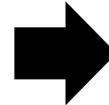
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Pseudomonas fluorescens + *Bacillus atrophaeus*

Arbuscular mycorrhizal fungi

horticulturae

Does Inoculation with Arbuscular Mycorrhizal Fungi Reduce Trunk Disease in Grapevine Rootstocks?

Taylor Holland¹, Patricia Bowen², Vasilis Kokkoris¹, Jose Ramon Urbez-Torres² and Miranda Hart^{1,*}

PeerJ

Commercial arbuscular mycorrhizal fungal inoculant failed to establish in a vineyard despite priority advantage

Corrina Thomsen¹, Laura Loverock¹, Vasilis Kokkoris^{3,4}, Taylor Holland¹, Patricia A. Bowen³ and Miranda Hart¹

agriculture

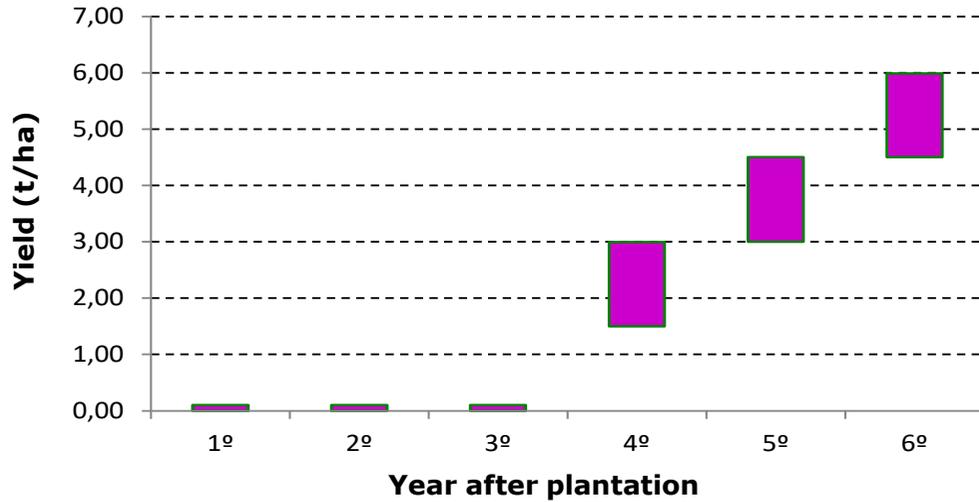
Performance and Establishment of a Commercial Mycorrhizal Inoculant in Viticulture

Daniel Rosa^{1,*}, Antreas Pogiatis¹, Pat Bowen², Vasilis Kokkoris³, Andrew Richards^{1,*}, Taylor Holland¹ and Miranda Hart¹

Commercial production settings

MANAGEMENT OF YOUNG VINEYARDS

Avoid placing a heavy fruit load on vines in the early production years



AVOID OVERCROPPING
Vines that are stressed are more susceptible to disease than unstressed vines.



2016
Plantation in 2014
Cultivar Macabeo

PHYTOPATHOLOGIA
MEDITERRANEA

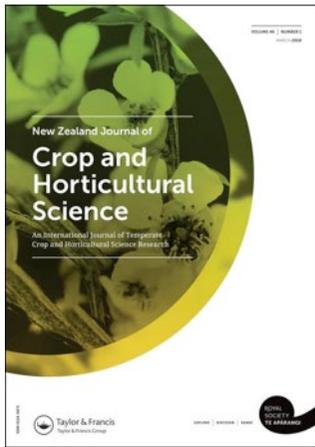
Grapevine trunk disease fungi: their roles as latent pathogens and stress factors that favour disease development and symptom expression

José Ramón ÚRBEZ-TORRES, Jared HRYCAN, Miranda HART, Patricia BOWEN, Thomas FORGE



3rd YEAR





New Zealand Journal of Crop and Horticultural Science

Publication details, including instructions for authors and subscription information:

<http://www.tandfonline.com/loi/tnzc20>

Grapevine propagation: principles and methods for the production of high-quality grapevine planting material

H Waite^{ab}, M Whitelaw-Weckert^{ac} & P Torley^{ab}

^a National Wine and Grape Industry Centre, Charles Sturt University, Wagga Wagga, Australia

^b School of Agricultural and Wine Sciences, Charles Sturt University, Wagga Wagga, Australia

^c New South Wales Department of Primary Industries, National Wine and Grape Industry Centre, Charles Sturt University, Wagga Wagga, Australia

Published online: 21 Nov 2014.

Managing Grapevine Trunk Diseases With Respect to Etiology and Epidemiology: Current Strategies and Future Prospects

David Gramaje¹

Instituto de Ciencias de la Vid y del Vino, Consejo Superior de Investigaciones Científicas - Universidad de la Rioja - Gobierno de la Rioja, Logroño 26007, Spain

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Mark R. Sosnowski

South Australian Research and Development Institute, GPO Box 397, Adelaide SA 5001, Australia; and School of Agriculture, Food and Wine, Waite Research Institute, The University of Adelaide, SA 5005, Australia

D. Gramaje, J. R. Úrbez-Torres, M. Sosnowski, 2018. Plant Dis. 102.

MOTHER FIELD

Rootstock



Scion



- Pruning wound protection: Chemicals and/or BCA
- Weed control
- Sanitation: removal of trimming debris
- Correct treatment and handling of harvested cuttings

NURSERY PROCESS

Hydration



- Cleaning of hydration tanks: frequently during the season, and at the start and end of the season
- Reduction of the cutting hydration period
- Application of Chemicals and/or BCA

Cold storage



- Cleaning of bins, boxes or crates before use in this phase
- Cleaning of cold storage room/s
- Application of Chemicals and/or BCA: as a dip for cuttings before storage

Disbudding



- Disinfection of pruning shears regularly
- Application of Chemicals and/or BCA: as a dip for cuttings after disbudding

Grafting



- HWT prior to grafting
- Disinfection of grafting machines regularly
- Application of Chemicals and/or BCA: as a dip for vines after grafting

Callusing



- Use moderate temperature for callusing and rooting
- Disinfection of callusing rooms regularly
- Application of Chemicals and/or BCA

NURSERY FIELD

Root and shoot development



- Application of Chemicals and/or BCA directly to soil
- Weed control

Uprooting and distribution



- Application of Chemicals and/or BCA: as a dip for one-year-old vines before storage as well as before despatch
- HWT of dormant nursery plants prior to despatch

Conclusions

- The **spread of fungal trunk pathogens** via infected nursery material is a long-established problem that cannot be solved quickly.
- Nurseries must undertake substantial efforts to **improve sanitation**.
- Tree crop nurseries are struggling to meet **demands for plant material**.

Almond nurseries in Spain

Phaeoacremonium minimum (20% incidence)



Marín-Terrazas et al. 2016 Plant Dis. 100

Blueberry nurseries in New Zealand



Four nurseries tested
positive (45% incidence)
Neofusicoccum spp.

Tennakoon et al. 2018 Eur. J. Plant Pathol. 150

Forest nurseries in Spain

17 nurseries
12 species
Cylindrodendrum
Dactylonectria
Ilyonectria



Mora-Sala et al. 2018 Plant Dis. 102

Olive nurseries in South Africa

2 nurseries
Nectriaceae
Diaporthaceae
Botryosphaeriaceae
Phaeomoniellaceae
Pleurotomaceae
Togniniaceae



Van Dyk et al. 2021 Plant Dis. (in press)

Can we really produce “disease-free” plant material even when following clean production best management practices?

CLEAN PLANT CERTIFICATION PROGRAM

Challenges

Broad range of taxonomically unrelated pathogens

Plant nursery conditions and practices favor fungal infection

Detection and identification requires destructive sampling

Develop fast and reliable methods for screening large number of plants for fungal trunk pathogens infections.

DNA-microarray (Úrbez-Torres et al. 2015 Phytopathology 105)

Use of NGS from asymptomatic tissues (Vettraino et al. 2017 Plos One 12)

Impact of global trade and implications of trunk disease infection in other tree crop nurseries.



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Thanks for your attention!

David Gramaje

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 **@biovitis**